Research Report

Potential contribution of oxidative stress and inflammation to anxiety and hypertension

Samina Salima,⁎ Mohammad Asghara, Manish Tanejaa, Iiriss Hovattab,c, Gaurav Chugha, Craig Vollerta, Anthony Vu

aDepartment of Pharmacological and Pharmaceutical Sciences, University of Houston, Houston, TX, USA
bResearch Programs Unit, Molecular Neurology and Department of Medical Genetics, Haartman Institute, University of Helsinki, Finland
cMental Health and Substance Abuse Services, National Institute for Health and Welfare, Helsinki, Finland

ARTICLE INFO

Article history:
Accepted 9 June 2011
Available online 16 June 2011

Keywords:
Oxidative stress
Anxiety
Antioxidants
Hypertension
Inflammation

ABSTRACT

Previously, we have published that pharmacological induction of oxidative stress causes anxiety-like behavior in rats and also is associated with hypertension in these animals. Here, we report that sub-chronic induction of oxidative stress via pharmacological induction leads to i) reduction in glyoxalase (GLO)-1 and glutathione reductase (GSR)-1 expression; ii) calpain mediated reduction of brain derived neurotrophic factor (BDNF) levels; iii) NFκB mediated upregulation of proinflammatory factors interleukin (IL)-6 and tumor necrosis factor (TNF)-α and elevated angiotensin (AT)-1 receptor levels in hippocampus, amygdala and locus coeruleus regions of the brain. Acute oxidative stress has opposite effects. We speculate that regulation of GLO1, GSR1, BDNF, NFκB and AT-1 receptor may contribute to anxiety-like behavior and hypertension in rats.

Published by Elsevier B.V.

1. Introduction

Recently, we and others have reported role of oxidative stress in anxiety-like behavior in rodents (de Oliveira et al., 2007; Hovatta et al., 2005; Masood et al., 2008; Salim et al., 2010a,b). Masood et al. (2008) reported that induction of oxidative stress in hypothalamus and amygdala occurs in parallel with anxiety in mice. Consumption of high sucrose diet was reported to increase protein oxidation in frontal cortex and induce anxiety in rats (Souza et al., 2007). Increased anxiety has been found to be positively correlated with increases in reactive oxygen species in granulocytes (Bouayed et al., 2007). In another study, oxidative stress in the adult rat hippocampus was reported to be anxiogenic (de Oliveira et al., 2007). Our own recent publications suggest that treatment with two separate oxidative stress inducers, L-Buthionine-(S,R)-sulfoximine (BSO) and xanthine plus xanthine oxidase (X+XO) both cause anxiety-like behavior (Salim et al., 2010a,b) and also increase blood pressure in rats (Salim et al., 2010b). Interestingly, induction of oxidative stress via a non-pharmacological method also leads to anxiety-like behavior in rats (Vollert et al., 2011). Despite a causal role of oxidative stress in anxiety (Salim et al., 2010a), as well as a positive correlation between oxidative stress and high blood pressure (Salim et al., 2010b), underlying neuronal mechanisms and specific molecules that potentially regulate anxiety and hypertensive behaviors remain unclear. In this

⁎ Corresponding author. Fax: +1 713 743 1229.
E-mail address: ssalim@uh.edu (S. Salim).

Abbreviations: X+XO, xanthine and xanthine oxidase; MDA, malondialdehyde; SDS-PAGE, sodium dodecyl sulfate polyacrylamide gel electrophoresis; OF, open-field; LD, light-dark

0006-8993/ $ – see front matter. Published by Elsevier B.V.
doi:10.1016/j.brainres.2011.06.024
study we provide novel mechanistic insights by revealing specific molecular targets that may be critical to the etiology of anxiety and hypertension, two of the highly prevalent comorbid diseases (McLaughlin et al., 2003; Roy-Byrne et al., 2008). By drawing attention to specific molecular targets, this study would open new avenues for research into the causal role of these molecules in anxiety and hypertension that may be amenable to pharmacological intervention.

Increasing evidence suggests a central role for oxidative and nitrosative stress elicited via generation of reactive oxygen species (ROS) and reactive nitrogen species (RNS), in a variety of congenital and acquired disorders of the central nervous system (CNS) (Calabrese et al., 2000; Halliwell, 2006; Sayre et al., 2008). This association is largely attributed to high vulnerability of brain to oxidative load (Ng et al., 2008). To cope with harmful effects of ROS and RNS, many detoxifying or antioxidant enzymes are expressed in the CNS (Ishii et al., 2000). Currently, two such enzymes have received great attention. Overexpression of glyoxalase (GLO)-1 and glutathione reductase (GR)-1 in mouse brain increased while inhibition of GLO1 expression decreased anxiety-like behavior of mice (Hovatta et al., 2005). A negative relationship between anxiety-like behavior and GLO1 expression has also been reported (Ditzen et al., 2006; Kromer et al., 2005; Landgraf et al., 2007; Thornalley, 2006). This disagreement is most likely due to genetic variation or different models studied (Williams et al., 2009). Regulation of GLO1 and GSR1 seems important for anxiety-like behavior no matter which studies we tend to consider. This is particularly intriguing considering the important role this enzyme system plays with regards to dicarbonyl glycation. Oxidative stress leads to formation of advanced glycation end-products (AGEs) (Thornalley, 2005) and contributes to cytotoxicity and inflammation. GLO–GSH system detoxifies dicarbonyl (Thornalley, 2003) and methylglyoxal (MG)-mediated cytotoxicity (Di Loreto et al., 2008). In connection to this, brain derived neurotrophic factor (BDNF) is protective against MG challenge, protects neurons from oxidative stress and promotes neurogenesis (Duman et al., 2008). An antioxidant role of BDNF has also been proposed (Chan et al., 2010) and BDNF is considered critical to the pathophysiology of anxiety disorders (Martinowich et al., 2007).

Relevant to this, increased intraneuronal calcium in the brain accompanies oxidative stress and activates calcium-activated proteases, the calpains (Ghosh and Greenberg, 1995; Kim et al., 2007). Ca²⁺/calmodulin protein kinase (CaMK)-IV is a substrate for calpain (Tremper-Wells and Vallano, 2005) and catalyzes phosphorylation of cAMP response element-binding protein (CREB) (Corcoran and Means, 2000). CREB is known to regulate gene expression of BDNF (Einat et al., 2003; Pandey, 2003). This pathway involving calpains, CAMKIV and CREB therefore seem important for regulation of BDNF. Moreover, activation of inflammatory pathways has been observed in patients with anxiety disorders (O’Donovan et al., 2010) as well as with hypertension (Saavedra et al., 2011). NFκB is activated upon oxidative stress and reported to intensify oxidative stress leading to inflammation and neuronal damage (Elks et al., 2009; Mattson and Camandola, 2001). NF-kappaB p50-deficient mice have been reported to show reduced anxiety-like behaviors (Kassed and Herkenham, 2004). Calpain dependent NFκB activation is also known (Shumway et al., 1999). NF-B binding sites are located on angiotensin II AT1 receptor promoter region (Cowell et al., 2005). Brain angiotensin system plays a significant role in cardiovascular control (Armando et al., 2003; Saavedra, 2005). In this study we provide a putative mechanism involving the above molecules that may be critical to the etiology of anxiety and hypertension and their comorbid phenotype.

2. Results

2.1. Differential regulation of GLO1 and GSR1 expression upon induction of acute and sub-chronic oxidative stress

GSR1 and GLO1 protein expression significantly increased after 3 day BSO treatment in the hippocampus (+150%: GSR1; +172%: GLO1), amygdala (+66%: GSR1; 67%: GLO1) and LC (+93%: GSR1; +61%: GLO1) (Fig. 1A, B) while their levels decreased after 7 day BSO treatment in hippocampus (-75%: GSR1; -58%: GLO1), amygdala (-85%: GSR1; -48%: GLO1) and LC (-44%: GSR1; -71%: GLO1) (Fig. 1C, D). GAPDH (loading control) remained unchanged. GSR1 and GLO1 levels were normalized in the tempol supplemented groups. GLO1 and GSR1 levels were not significantly altered in cerebellum, cortex, striatum and hypothalamus (data not shown). Tempol treatment alone did not alter GSR1 or GLO1 at 3 or 7 day BSO (Fig. 1E). X+XO treatment for 7 days significantly decreased levels of GLO1 and GSR1 proteins in the hippocampus (-50%: GSR1; -60%: GSR1), amygdala (-72%: GLO1; -53%: GSR1) and LC (-73%: GLO1; -71%: GSR1) as compared to the vehicle treated control samples. GAPDH remained unchanged (Fig. 2).

2.2. Induction of sub-chronic oxidative stress activates calpain pathway and decreases CAMKIV, CREB and BDNF protein expression

X+XO treatment for 7 days significantly increased micro (μ) and millimolar (m)-calpain expression (indicators of calpain activation) as compared to the vehicle treated control samples of tissue homogenates prepared from the hippocampus (+98%: μ-calpain; +91% m-calpain), amygdala (+100%: μ-calpain; +140% m-calpain) and LC (+102%: μ-calpain; +101% m-calpain) (Fig. 3A, B). X+XO treatment significantly decreased levels of CAMKIV (Fig. 3C), P-CREB/total CREB (Fig. 3D) and BDNF (Fig. 3E) proteins in the hippocampus (-26%: CAMKIV; -75%: p/totalCREB, -100%: BDNF) amygdala (-57%: CAMKIV; -49%: p/total CREB, -65%: BDNF) and LC (-49%: CAMKIV; -72%: p/total CREB, -51%: BDNF) as compared to the controls. Levels of BDNF were increased with 3 day BSO treatment in hippocampus (+135%), amygdala (+211%) and LC (+309%) (Fig. 4A) while 7 day BSO decreased BDNF in the hippocampus (-61%), amygdala (-74%) and LC (-53%) as compared to the vehicle treated controls (Fig. 4B). GAPDH remained unchanged.

2.3. Induction of sub-chronic oxidative stress activates NFκB and increases AT1 receptor expression

X+XO treatment significantly increased levels of NFκB in the nuclear extracts prepared from hippocampus (+69%), amygdala (+111%) and LC (+81%) (Fig. 5A). This treatment increased AT1
receptor expression (Fig. 5B) in the hippocampus (+107%), amygdala (+62%) and LC homogenates (+77%) as compared to the vehicle treated controls. Lamin B and GAPDH used as a loading control for NFκB and AT1 receptor respectively remained unchanged.

2.4. Induction of sub-chronic oxidative stress increases pro-inflammatory markers

The levels of CRP and IL-6, markers of inflammation, in the plasma increased in rats treated with X+XO (+93%; CRP; +54%:

Fig. 1 – Effect of 3 and 7 day BSO treatment on GLO1 and GSR1 protein expression in hippocampus (HIP), amygdala (AMY) and locus coeruleus (LC). Tissue homogenates (15–25 μg proteins) were subjected to western blotting using GLO1, GSR1 and GAPDH (loading control) antibodies. Panels A and B indicate 3 day BSO treatment while panels C and D indicate 7 day BSO treatment. Left hand side of panel E indicates 3 day BSO treatment while right hand side shows 7 day BSO treatment. Lane 1: control; 2: BSO; 3: Tempol+BSO. Protein samples in C and D were loaded on a standard 10-well 8–16% SDS-PAGE gel while samples in A, B and E were loaded on a 48-well E-PAGE™ 48 Protein electrophoresis System (Invitrogen Corp. #EP048-08). E: western blotting for GLO1 and GSR1 indicate no change in controls, vehicle: C and tempol alone: T. H: hippocampus, A: amygdala, L: locus coeruleus.

*Significantly different from control, # from BSO p<0.05, ANOVA, N=12 rats.

Fig. 2 – Effect of X+XO treatment on GLO1 and GSR1 protein expression in hippocampus (HIP), amygdala (AMY) and locus coeruleus (LC). Tissue homogenates (25 μg proteins) were subjected to western blotting, using GLO1, GSR1 and GAPDH (loading control) antibodies. Shown are representative blots and densitometric ratios of proteins normalized to GAPDH, respectively.

*Significantly different from control, # from BSO p<0.05, ANOVA, N=6 rats.
IL-6) (Fig. 6A, B) or with 7 day BSO (+97%: CRP; +60%: IL-6) when compared to their controls. Tempol treated rats had CRP and IL-6 comparable to controls (Fig. 6C, D). Levels of TNF-α were also evaluated. TNF-α significantly increased in hippocampus (+103%), amygdala (+107%) and LC (+47%) in X+XO rats as compared to controls (Fig. 7A). TNF-α levels also increased in hippocampus (+137%), amygdala (+95%) and LC (+71%) in 7 day BSO treated rats. Tempol treatment normalized TNF-α (Fig. 7B).

3. Discussion

Recently, we published two studies reporting behavioral consequences of pharmacological induction of oxidative stress. In one, we established that oxidative stress produced via sub-chronic BSO treatment (7 days) caused anxiety-like behavior in rats while acute BSO treatment (3 days) did not (Salim et al., 2010a). In another study we demonstrated that oxidative stress produced via X+XO treatment caused anxiety-like behavior in rats. This treatment also was associated with hypertension in these animals (Salim et al., 2010b). Here we provide some biochemical results using blood and brain tissue homogenates from the rats utilized in the above studies.

In the present study we report that GLO1, GSR1 and BDNF levels were differentially regulated upon acute versus sub-chronic oxidative stress with BSO or X+XO treatments. Reduced levels of GLO1, GSR1 and BDNF were observed in hippocampus, amygdala and LC upon sub-chronic induction of oxidative stress with BSO or with X+XO treatments. Interestingly, GLO1, GSR1 and BDNF protein expression increased in these brain areas upon acute treatment. It seems that amygdala, LC and hippocampus are somewhat equally responsive to changes in oxidative stress (Salim et al., 2010a,b). Involvement of these brain areas in anxiety disorders is known (Charney and Drevets, 2002; Dell’Osso et al., 2010; Lopez et al., 1999; Shin et al., 2006) but their exact role in regulation of anxiety remains unclear (Menard and Treit, 1999, Shin et al., 2006). Although there is enough consensus in the field regarding the role of amygdala in anxiety (Menard and Treit, 1999), involvement of other brain regions is still a matter of debate (Davis, 2006; Maren, 2008; Quirk and Mueller, 2008). It is believed that gamma amino butyric acid (GABA) projections transmit anxiety-related information from the amygdala to various centers in the brainstem, including, LC (Dong et al., 2001; LeDoux et al., 1988) and LC provides primary noradrenergic projections to the hippocampus and amygdala. Thus a critical inter-connection made within these regions may be critical for regulation of oxidative stress-mediated anxiety. It is clear that high anxiety (Salim et al., 2010a,b) corresponds to low GSR1, GLO1 and BDNF while no anxiety (Salim et al., 2010a) with high levels of these proteins in these models. In agreement with these findings, our recent publication using a neuronal cell culture model of acute oxidative stress, suggests that GLO1 and GSR1 are transcriptionally up-
regulated via Regulator of G-protein Signaling protein (RGS)-2 dependent mechanisms (Salim et al., 2011). Sub-chronic oxidative stress in this cell line reduced RGS2, GLO1 and GSR1 mRNA and proteins (unpublished observations).

Furthermore, X+XO mediated sub-chronic induction of oxidative stress increased calpain expression in the hippocampus, amygdala and LC, accompanied with a concomitant reduction in CAMKIV and CREB levels. CAMKIV is a substrate of calpain and regulates CREB levels which in turn modulate BDNF expression (Einat et al., 2003; Pandey, 2003). Thus BDNF levels are down-regulated perhaps due to calpain dependent degradation of its upstream regulators, CAMKIV and CREB. These are interesting observations considering that induction of oxidative stress causes protein glycation and GLO-GSH system detoxifies dicarbonylation (Thornalley, 2003). And, BDNF is reported to be upregulated upon MG challenge. Perhaps BDNF levels increase during acute oxidative stress (3 days BSO) as an immediate protective response against glycation. As a result of this compensatory response antioxidant defense (GLO1 and GSR1 expression) also is upregulated. Upon chronic oxidative stress (7 days BSO or 7 days X+XO) this adaptive mechanism is lacking due to calpain dependent degradation of BDNF. It is

Fig. 4 – Effect of 3 and 7 days of BSO treatment on BDNF protein expression in hippocampus (HIP), amygdala (AMY) and locus coeruleus (LC). Tissue homogenates (25 μg proteins) were subjected to western blotting, using anti-BDNF and GAPDH (loading control) antibodies. Panel A indicate 3 day BSO treatment while panel B indicates 7 day BSO treatment. Shown are representative blots and densitometric ratios of proteins normalized to GAPDH, respectively. *Significantly different from control, # from BSO (B) p<0.05, ANOVA, N=6 rats.

Fig. 5 – Effect of X+XO treatment on NFκB and AT1 receptor protein levels in hippocampus (HIP), amygdala (AMY) and locus coeruleus (LC). Tissue homogenates (25 μg proteins) or nuclear extracts of different brain areas were subjected to western blotting, using anti-NFκB, AT1, lamin B (loading control for nuclear proteins) and GAPDH (loading control) antibodies. Shown are representative blots and densitometric ratios of proteins normalized to lamin B or GAPDH, respectively. *Significantly different from control, # from X+XO p<0.05, ANOVA, N=6 rats.
reasonable to suggest that decline in GLO1 and GSR1 leads to excessive protein glycation. BDNF can no longer compensate for this defect and impairment in this detoxification mechanism occur inducing irreversible alterations in the antioxidant homeostasis in the brain. Moreover, BDNF is not only considered as protective against oxidative stress but is also known to promote neurogenesis (Duman et al., 1997) and synaptic plasticity (Schaaf et al., 2001). Thus it raises an intriguing possibility that decreased BDNF favors a condition reflective of high oxidative stress and reduced plasticity, all leading to a potentially high anxiety state.

Furthermore, our results demonstrate increased levels of transcription factor, NFκB in nuclear extracts of hippocampus, amygdala and LC samples obtained from X+XO treated rats, accompanied with increased angiotensin II AT1 receptor protein levels. Calpain dependent NFκB activation has been reported (Shumway et al., 1999). NFκB binding sites are located on the AT1 receptor promoter region (Cowling et al., 2005). Therefore, we postulate that oxidative stress activates calpains, which via NFκB lead to transcriptional activation of AT1 receptor thus increasing AT1 expression. Role of amygdala, LC and hippocampus is proposed in central control of blood pressure (Elam

**Fig. 6** – Effect of 7 day X+XO and BSO treatment on plasma levels of CRP and IL-6 in rats. CRP and IL-6, markers of inflammation were examined using kit based ELISA assays in rat plasma of control, X+XO and BSO-treated rats. *Significantly different from control rats p<0.05, ANOVA, N=6–10 rats.

**Fig. 7** – Effect of X+XO or BSO treatment on TNF-α in brain homogenates of rats. Brain homogenates were subjected to ELISA for measurement of TNF-α levels according to the manufacturer’s protocol (R&D Systems, Minneapolis, MN). *Significantly different from control rats p<0.05, ANOVA, N=6–8 rats.
et al., 1985; Pietranera et al., 2008; Saha, 2005) but information regarding precise neural mechanisms operated within these regions that enable such control is lacking. Which out of the three regions is lesser, equal or more effective than the other in regulation of hypertension via oxidative stress mechanisms is an interesting topic for future studies. Many studies have linked central angiotensin activity with elevated blood pressure. In fact, blockade of the brain angiotensin II AT1 receptors contribute to blood pressure decrease in hypertension, in several animal models (Armando et al., 2003; Saavedra, 2005). Furthermore, NFκB activation is reported to promote inflammation leading to neuronal damage (Elks et al., 2009; Mattson and Camandola, 2001). Increased pro-inflammatory cytokines including interleukin-6 (IL-6) and TNF-α have been detected in anxiety disorder (O’Donovan et al., 2010) and hypertensive patients (Saavedra et al., 2011). Our results are in agreement with these reports. We observed increased CRP-1 and IL-6 in the serum and elevated TNF-α in the brain of 7 days X+XO as well as 7 day BSO treated rats.

Taken together, our observations have prompted the following speculation (Fig. 8). Sub-chronic oxidative stress i) decreases GLO1 and GSR1 increasing dicarbonyl glycation; ii) activates calpain to degrade BDNF causing detoxification impairment, contributing to greater oxidative stress, reducing neurogenesis and plasticity; iv) promotes NFκB-dependent inflammation; all of these factors contribute to co-morbidity of anxiety-like behavior and hypertension in rats.

4. Experimental procedures

4.1. Animal model of oxidative stress

All experiments were conducted in accordance with the NIH guidelines using approved protocols from the University of Houston Animal Care and Use Committee. Male Sprague–Dawley rats (200–250 g) 7–8 weeks of age were utilized, oxidative stress produced as described (Salim et al., 2010a,b) and brains harvested as below.

4.2. Brain dissections and preparation of homogenates

After conducting anxiety-like behavior tests (Salim et al., 2010a,b), rats were anesthetized, brains removed, frozen and brain stem removed (Salim et al., 2010b). Hippocampus, amygdala and locus coeruleus (LC) were identified according to Paxinos and Watson (1986) and isolated as before (Salim et al., 2010a,b). Homogenates were prepared as published (Salim et al., 2010a,b).

4.3. Western blot analysis

Homogenates were subjected to SDS-polyacrylamide gel electrophoresis (PAGE) and western blotting using primary (GLO1, GSR1, calpains, BDNF, CAMKIV, phospho and total CREB, NFκB, AT1 receptor, lamin B and GAPDH) (overnight, 4 °C) and horseradish peroxidase (HRP)-conjugated secondary (room temperature, 1 h) antibodies. Band intensities were determined on Alpha Ease FC 4.0 (Alpha Innotech Corp., San Leandro, CA).

4.4. C-reactive protein (CRP) measurement

C-reactive protein is an acute phase protein, the levels of which increase in inflammation and is considered as a marker of inflammation (Schaap et al., 2006). CRP was measured by CRP-ELISA kit using purified rat CRP as standard supplied with the kit (Immunology Consultants Labs., Inc., Newberg, OR).

4.5. Interleukin-6 (IL-6) and TNF-α measurement

Rat plasma was coated on a 96-well plate (overnight, 4 °C), washed, incubated with anti-rat IL-6 primary (cat #ARCO062, Biosource, Camarillo, CA) and HRP-linked secondary antibody and the color developed with tetramethyl benzidine (Substrate System for ELISA, Sigma Chem. Co., St. Louis, MO) and 1 N H2SO4 was read at 450 nm (ELISA reader; BIO-TEK Instruments, Inc., Winooski, Vermont, USA) according to the manufacturer’s protocol. TNF-α measurement was done using ELISA kit as recommended (R&D Systems, Minneapolis, MN).

4.6. Data analysis

Data are expressed as mean±SEM. Significance was determined by one or two way ANOVA as appropriate by applying Tukey’s post-hoc test (GraphPad Software, Inc. San Diego, CA). A value of p<0.05 was considered significant.
Acknowledgments

Funding: GEAR grant, UH (S.S), SURF award (CV), PURS award (AV), NIH/NIAAG29904 (MA) and Academy of Finland NEURO research program and Academy Research Fellowship (IH).

REFERENCES


mineralocorticoid-induced hypertension


Roy-Byrne, P.P., Davidson, K.W., Kessler, R.C., Asmundson, G.J.,
Goodwin, R.D., Kubzansky, L., Lydiard, R.B., Massie, M.J., Katon,
W., Laden, S.K., Stein, M.B., 2008. Anxiety disorders and

Saavedra, J.M., 2005. Brain angiotensin II: new developments,
unanswered questions and therapeutic opportunities. Cell.

brain angiotensin II AT1 receptors ameliorates stress, anxiety,
brain inflammation and ischemia: therapeutic implications.
Psychoneuroendocrinology (1), 1–18.

Saha, S., 2005. Role of the central nucleus of the amygdala in the
control of blood pressure: descending pathways to medullary

Salim, S., Sarraj, N., Taneja, M., Saha, K., Tejada-Simon, M.V., Chugh,
C., 2010a. Moderate treadmill exercise prevents oxidative

Salim, S., Asghar, M., Chugh, G., Taneja, M., Xia, Z., Saha, K., 2010b.
Oxidative stress: a potential recipe for anxiety, hypertension

Salim, S., Asghar, M., Taneja, M., Hovatta, I., Wu, Y.L., Saha, K., Sarraj,
homeostasis in neuronal cells. FEBS Lett. 585 (9), 1375–1381 May 6
(Electronic publication ahead of print, 2011 Apr 16).

Sayre, L.M., Perry, G., Smith, M.A., 2008. Oxidative stress and

Correlation between hippocampal BDNF mRNA expression and

markers and loss of muscle mass (sarcopenia) and strength. Am.

prefrontal cortex, and hippocampal function in PTSD. Ann.
N. Y. Acad. Sci. 1071, 67–79.

Shumway, S.D., Maki, M., Miyamoto, S., 1999. The PEST domain
of IkB is necessary and sufficient for in vitro degradation by

Souza, C.G., Moreira, J.D., Siqueira, I.R., Pereira, A.G., Rieger, D.K.,
palatable diet consumption increases protein oxidation in rat
frontal cortex and anxiety-like behavior. Life Sci. 81 (3), 198–203
Jun 27 (Electronic publication ahead of print, 2007 May 17).

Thornalley, P.J., 2003. Glyoxalase I structure, function and a critical
role in the enzymatic defence against glycation. Biochem. Soc.
Trans. (6), 1343–1348.

Thornalley, P.J., 2005. Dicarbonyl intermediates in the Maillard

Thornalley, P.J., 2006. Unease on the role of glyoxalase 1 in

Ca2+-dependent signaling via proteolysis of nuclear Ca2+/
calmodulin-dependent protein kinase type IV in cultured

Vollert, C., Zagaar, M., Hovatta, I., Taneja, M., Vu, A., Dao, A.,
Levine, A., Alkadhi, K., Salim, S., 2011. Exercise prevents sleep
derivation-associated anxiety-like behavior in rats: potential
role of oxidative stress mechanisms. Behav. Brain Res. May 19.
[Epub ahead of print].

Williams IV, R., Lim, J.E., Harr, B., Wing, C., Walters, R., Distler,
M.G., Teschke, M., Wu, C., Tarantino, L.M., Borevitz, J.O.,
Palmer, A.A., 2009. A common and unstable copy number
variant is associated with differences in Glo1 expression and